

# An *In Vivo* Therapeutic Antibody Efficacy Evaluation: the Chicken Embryo's CAM-based Assay

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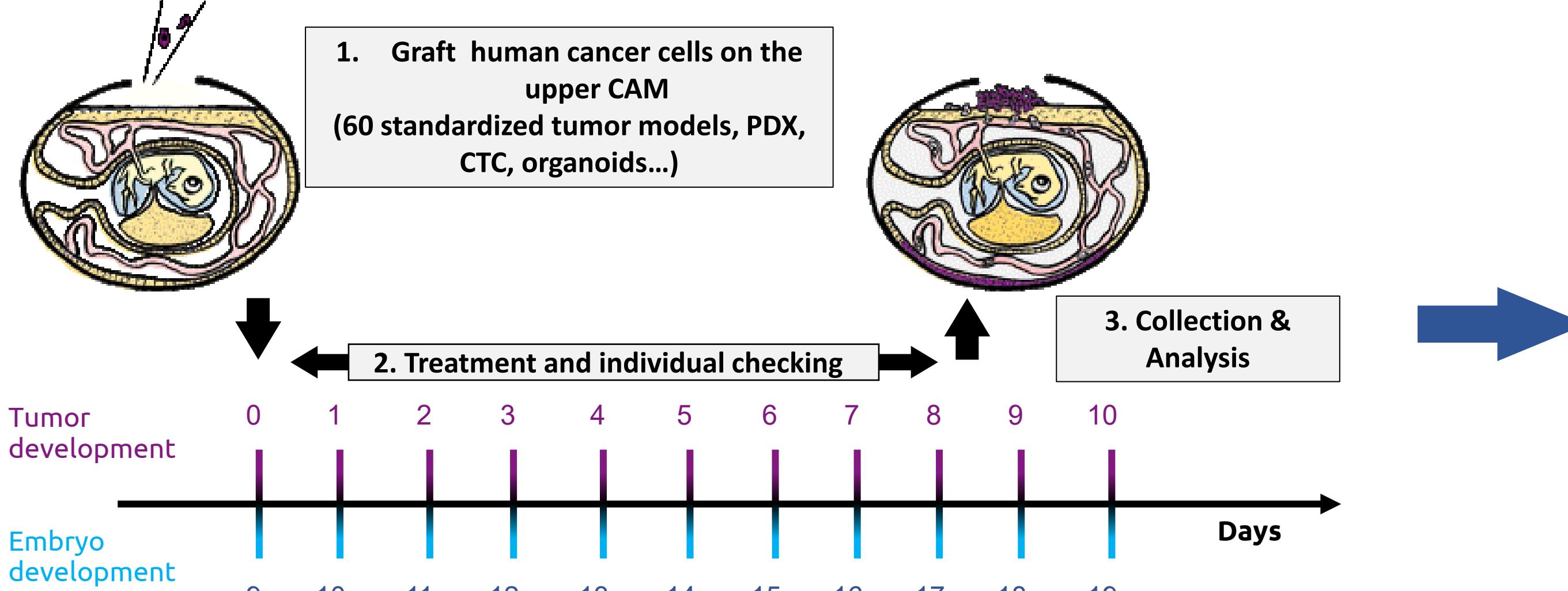
# in9votion

WHICH MOLECULE WILL STAND OUT

## INTRODUCTION

For cancer drug development, a pertinent preclinical model is essential for the rapid and efficient transition from preclinical evaluation to clinical progress. Up until now, rodents are the most-often used models for preclinical evaluation. However, their use presents several drawbacks, including ethical constraints (3Rs), time-consuming and costly experiments, and the immunodeficiency of humanized models, which could slow down drug development [1, 2]. Since their introduction, xenografts on the chicken embryo's ChorioAllantoic Membrane (CAM) have proven extremely valuable for *in vivo* studies in cancerology [3]. They are suitable for studying tumor development, angiogenesis, malignant cell dissemination, and for estimating the toxicity and the efficacy of novel therapies [4]. Here, we demonstrate that the CAM-based *in ovo* model, which has an active immune system, is useful for rapidly testing and comparing therapeutic antibodies' efficacy, such as anti-PD1/PDL1 Abs, Antibody Drug Conjugates (ADCs), and others, on tumor growth and metastatic invasion. Our results clearly show the power of the chicken embryo model as an efficient *in vivo* model for testing a large spectrum of cancer drugs.

## MATERIALS AND METHODS

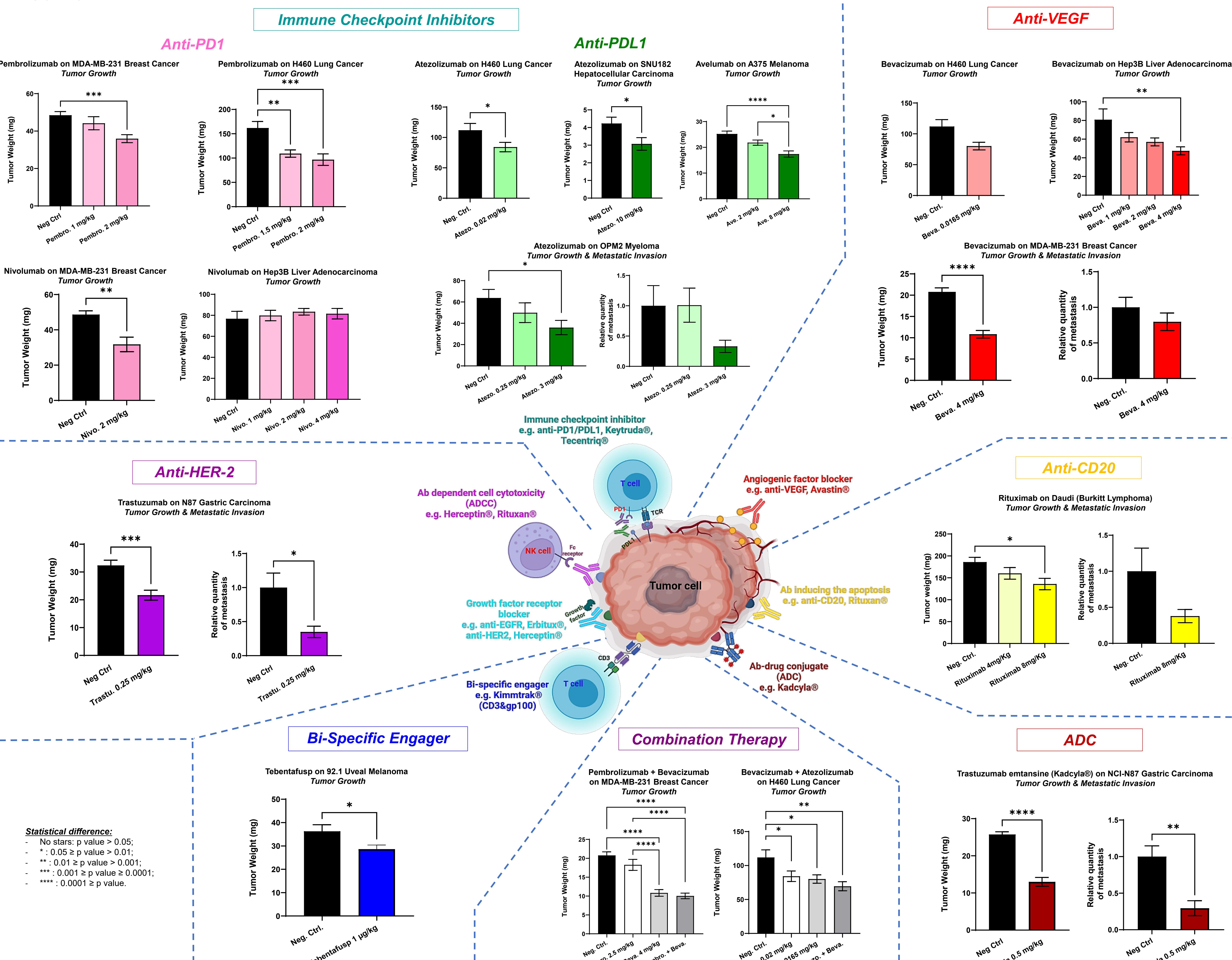


### Available *in ovo* readouts:

- Drug toxicity (embryo survival rate & malformation tracking)
- Tumor growth (tumor weight)
- Metastatic invasion in the lower CAM (away from the tumor), and in embryonic tissues
- Immune cell infiltration (RT-qPCR analysis of intra-tumoral immune cell biomarkers)
- Angiogenesis
- Xenografted tumor morphology: histological and immunohistochemistry (IHC) analysis
- Transcriptomic analysis of the tumor for exploring mechanisms of action

NB: This list is non exhaustive.

## RESULTS



## CONCLUSIONS

The chicken embryo is an immune competent, *in vivo* model, and a robust immune response can be induced *in ovo* [5]. Our results show that the CAM model is suitable for testing therapeutic antibodies, immune therapies and ADCs *in vivo*. The presence of an active immune system allows the characterization of antibodies' effects on tumor cells, the tumor microenvironment, and the entire organism. These findings demonstrate both the pertinent use of this model for testing therapeutic antibodies, and also its high potential for testing more cancer drugs on a large spectrum, as well for validating combination regimens. Furthermore, many additional analyses can also be considered using the CAM assay, including neo-angiogenesis analysis, metastatic invasion, transcriptomics, and many others. More generally, the chicken embryo model has shown many strong advantages over classical models, including the simplicity of egg management and handling, cost-effectiveness, time efficiency, and 3Rs compliance (it is not considered to be an animal model). All these benefits illustrate that the chicken embryo model is a viable, alternative *in vivo* model, which is fast and reliable for use in cancer drug discovery.

## References

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